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14. ABSTRACT

One of the key questions in Paget's disease of bone (PDB) is the nature of the "trigger" for initiation of the disease. Inheritance of a predisposing mutation and childhood infection with Measles virus are important, but apparently insufficient to initiate the disease until a significant period of time has passed. Understanding what occurs in this intervening time period is the goal of this proposal. Our model involves the opposing action of two Measles virus genes. The MVV gene suppresses gene expression as part of the virus' ability to promote persistent infection. The MVNP gene appears to activate gene expression to promote acute infection. In our model, when Measles virus infects a bone cell, the MVV gene creates a latent infection by suppressing gene expression in the cell. Then, a chance genetic event results in loss of the MVV gene and unmasking the MVNP gene, which then cooperates with the mutated SQSTM1 to initiate the exaggerated pattern of bone cell growth characteristic of PDB. We are presently testing this model. We have had difficulty cloning the MVV gene but propose a number of strategies to overcome this problem.

15. SUBJECT TERMS

Paget's disease of bone, Measles virus, osteoclast, osteoblast, disease initiation, latent viral infection, Sequestosome 1, Measles V gene, Measles Nucleocapsid gene

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Table of Contents

	-	<u>Page</u>
1.	Introduction	4
2.	Keywords	4
3.	Overall Project Summary	. 4
4.	Key Research Accomplishments	7
5.	Conclusion	. 7
6.	Publications, Abstracts, and Presentations	. 7
7.	Inventions, Patents and Licenses	. 7
8.	Reportable Outcomes	. 7
9.	Other Achievements	7
10). References	7
11	. Appendices	8

Introduction

The long-term goal of this project is to understand how Paget's Disease of Bone (PDB) begins. PDB can be inherited in families and several genes have been linked to PDB including Sequestosome 1 (SQSTM1), which has been linked to 40% of familial PDB¹. Measles virus infection has also been linked to PDB in a substantial fraction of cases^{2,3}. One of the key questions in PDB is the nature of the "trigger" for initiation of the disease⁴. As an illustrative example, individuals in a family with familial PDB inherit a germline mutation in the SOSTM1 gene that predisposes those individuals to PDB. A predisposed individual contracts measles at age 8, which is the only other known factor for PDB. However, the disease does not reveal itself until that individual reaches age 50. What has occurred in the intervening time period that was not present at age 8 is the basis for this proposal. Our model involves the measles virus. The measles virus genome contains a number of genes. Two of these genes, the MVNP protein and the MVV gene appear to act in opposing fashion to control gene expression in an infected cell^{2,5}-9. The MVV gene suppresses gene expression as part of the virus' ability to evade immunological surveillance and promote persistent infection^{5,6}. In contrast, the MVNP gene appears to activate gene expression to promote acute infection¹⁰⁻¹². Our model is that when the measles virus infects a bone cell, the MVV gene creates a latent infection state by suppressing gene expression in the cell. Then, over the years, a chance genetic event in a single bone cell containing the measles virus results in the loss of the MVV gene. At this time, the MVNP gene is unmasked and cooperates with the mutated SQSTM1 to initiate the exaggerated pattern of bone cell growth characteristic of PDB. To test this model, we proposed two specific aims. In the first aim, we proposed to introduce both the MVV and MVNP genes into bone cells in culture and then use siRNA technology to turn off the MVV gene and observe the change in the growing bone cells to see if it mimics PDB. In the second aim, we proposed to examine affected bone tissue from patients with PDB to see if the affected bone has evidence of mutation of the MVV gene while maintaining the MVNP gene. If our hypothesis is true, then this will dramatically alter our understanding of how PDB initiates. It will also enlighten our understanding of the relationship between measles and PDB.

Keywords

Paget's disease of bone, Measles virus, osteoclast, osteoblast, disease initiation, latent viral infection, *Sequestosome 1*, V gene, Nucleocapsid gene

Overall Project Summary

We have requested a No-Cost Extension to complete the work proposed in this application. We believe that the unexpected difficulties that we encountered during the funding year can be overcome and the work completed with an extension of time. Thus in addition to the description of the work done in the last year, we also propose solutions that we think can overcome the problems.

There were two specific aims to the proposal. The first specific aim was to determine whether loss of expression of Measles virus V (MVV) gene in the presence of Measles virus NP (MVNP) gene leads to a pagetic phenotype. In this aim there were three tasks. The first task was to clone the open reading frame of the MVV gene and the MVNP gene in such a fashion as to allow the two genes to be co-transfected and detected using different fluorescent tags. Our plan was to compare the expression of the MVV gene with the expression of the MVNP gene, which is critical for the onset of Paget's disease. However, the MVV gene is a part of the measles virus

genome in which all three reading frames encode distinct genes (MVP, MVV and MVC) using the same sequence. Our problem has been to clone and express the MVV gene, which contains a non-templated RNA edit. Thus far, we have been unsuccessful.

The problem is shown in Figure 1. The MVP and MVV genes are encoded by a common sequence in the measles virus genome in which both use the ATG start site at nucleotide 1807 and from 1807 to 2498 show a common translated amino acid sequence. The two sequences diverge due to a non-templated insertion of a G nucleotide at nucleotide 2499 in the MVV sequence resulting in a frameshift and altered reading resulting in a unique amino acid sequence for MVP and MVV from that point on. Our attempts at cloning this sequence of the MVV gene from the Measles' genome using a PCR-based strategy in which we assemble the MVV gene from the measles virus genomic RNA using overlapping primer sets have not been successful. As a result, what was expected to be completed in the first six months of the proposal has taken the complete year and still not been successful. We have two proposed solutions to this problem. The first is to adopt a two-step procedure where a truncated MVP gene sequence (approximately nucleotides 1800 - 3000) is amplified by PCR and cloned into a vector and then the nontemplated MVV G nucleotide at 2499 is inserted into the cloned MVP gene sequence by a second PCR reaction or restriction reaction. This may reduce the problem of trying to insert a non-templated G directly into the RNA sequence from the total Measles genome since we will have increased the quantity of template we can use. Our second alternative is to isolate the MVV RNA from cells infected with the intact Measles virus and then use that RNA as template to clone the MVV gene. This has the advantage of letting the virus insert the non-templated G base into the RNA sequence. However, this is our second choice because we had been attempting to avoid having intact Measles virus in the laboratory but this may be our only solution to the problem. Once the MVV RNA is isolated, we can use it as a template to reverse transcribe it and clone it into a suitable vector. In either case, we can then transfect the cloned MVV gene together with the MVNP gene. Specific Aim 1 tasks II and III can than be accomplished once the MVV and MVNP genes are co-expressed in the appropriate cells.

Specific Aim 2 was to screen matched normal and pagetic bone samples for evidence of MVV and MVNP and demonstrate that the MVV present in the pagetic samples is mutated in a way that causes inactivation of the MVV gene product. Here we had difficulty with the qRT PCR amplification of the MVV gene. We were not able to detect amplification of the MVV genes in the formalin-fixed paraffin-embedded (FFPE) tissue samples. This is possibly due to our choice of primers, which needed to distinguish MVP from MVV, resulting in a primer sequence location that was not optimal for qRT PCR. Without a cloned MVV gene as a positive control, it is difficult to determine what went wrong. As an alternative approach, we propose to try a ligand-mediated amplification-based assay, which should allow us to distinguish MVP from MVV by the difference in the non-templated inserted G that distinguishes MVP from MVV. We also anticipate that a successful cloning of the MVV gene and subsequent transfection of a control cell line will allow us to troubleshoot the process and succeed in amplifying and detecting the MVV gene in the FFPE tissues. At this point, the amplified gene products can be sequenced to complete our Specific Aim 2 tasks.

Personnel receiving pay for research effort were Marc F. Hansen, Principal Investigator; Michael Mogass, Postdoctoral Fellow; Cindy Alander, Research Assistant.

1807	ATGqcaqaaqaqcaqqcacqccATGtcaaaaaacqqactqqaatqtatccqqqctctcaaq				
100,	M A E E Q A R H V K N G L E C I R A L K	MVP gene			
	M A E E Q A R H V K N G L E C I R A L K	MVV gene			
	M S K T D W N V S G L S R	-			
1867	gccgagcccatcggctcgctggccgtcgaggaagccatggcagcatggtcagaaatatca	-			
	A E P I G S L A V E E A M A A W S E I S	MVP gene			
	A E P I G S L A V E E A M A A W S E I S	MVV gene			
1005	P S P S A R W P S R K P W Q H G Q K Y Q	MVC gene			
1927	gacaacccaggacaggaccgagccacctgcaaggaaaggaagg	MITD cono			
	D N P G Q D R A T C K E E K A G S S G L D N P G Q D R A T C K E E K A G S S G L	MVP gene MVV gene			
	T T Q D R T E P P A R K R R Q A V R V S	MVC gene			
1987	agcaaaccatgcctctcagcaattggatcaactgaaggcggtgcacctcgcatccgcggt	3			
	S K P C L S A I G S T E G G A P R I R G	MVP gene			
	S K P C L S A I G S T E G G A P R I R G	MVV gene			
0045	ANHASQQLDQLKAVHLASAV	MVC gene			
2047	cagggatctggagaggcgatgacgacgctgaaactttgggaatcccctcaagaaatctc	MITD cono			
	Q G S G E S D D D A E T L G I P S R N L Q G S G E S D D D A E T L G I P S R N L	MVP gene MVV gene			
	R D L E R A M T T L K L W E S P Q E I S	MVC gene			
2107	caggcatcaagcactgggctacagtgttatcatgtttatgatcacagcggtgaagcggtt	3			
	Q A S S T G L Q C Y H V Y D H S G E A V	MVP gene			
	Q A S S T G L Q C Y H V Y D H S G E A V	MVV gene			
	R H Q A L G Y S V I M F M I T A V K R L	MVC gene			
2167	aagggaatccaagatgctgactctatcatggttcaatcaggccttgatggtgatagcacc	MID cone			
	K G I Q D A D S I M V Q S G L D G D S T K G I Q D A D S I M V Q S G L D G D S T	MVP gene MVV gene			
	RESKMLTLSWFNQALMVIAP	MVC gene			
2227	ctctcaggaggagacgatgaatctgaaaacagcgatgtggatattggcgaacctgatacc	9			
	LSGGDDESENSDVDIGEPDT	MVP gene			
		MVV gene			
	S Q E E T M N L K T A M W I L A N L I P	MVC gene			
2287	gagggatatgctatcactgaccggggatctgctcccatctctatggggttcagggcttct				
	E G Y A I T D R G S A P I S M G F R A S	MVP gene			
	E G Y A I T D R G S A P I S M G F R A S R D M L S L T G D L L P S L W G S G L L	MVV gene MVC gene			
2347	qatqttqaaactqcaqaaqqtqaqatccacqaqctcctqaqactccaatccaqaqqc	nvc gene			
2017	D V E T A E G G E I H E L L R L Q S R G	MVP gene			
	D V E T A E G G E I H E L L R L Q S R G	MVV gene			
	M L K L Q K E V R S T S S -	MVC gene			
2407	aacaactttccgaagcttgggaaaactctcaatgttcctccgcccccgaaccccggtagg				
	N N F P K L G K T L N V P P P P N P G R	MVP gene			
2467	N N F P K L G K T L N V P P P P N P G R gccagcgcttccgagacacccattaaaaag gc cagacgcgagattagcctcatttgga	MVV gene			
2407	A S A S E T P I K K T D A R L A S F G	MVP gene			
	A S A S E T P I K K G H R R E I S L I W	MVV gene			
2527	acggagatcgcgtctttattgacaggtggtgcaacccaatgtgctcgaaagtcaccctcg	_			
	T E I A S L L T G G A T Q C A R K S P S	MVP gene			
	N G D R V F I D R W C N P M C S K V T L	MVV gene			
2587	gaaccatcagggccaggtgcacctgtggggaatgtccccgagtgtgtgagcaatgccgca	MITTO			
	E P S G P G A P V G N V P E C V S N A A G T I R A R C T C G E C P R V C E Q C R	MVP gene MVV gene			
2647	ctgatacaggagtggacacccgaatctggtaccacaatctccccgagatcccagaataat	nvv gene			
	L I Q E W T P E S G T T I S P R S Q N N	MVP gene			
	T D T G V D T R I W Y H N L P E I P E -	MVV gene			
2707	$\tt gaagaagggggagactattatgatgatgatgttctccgatgtccaagacatcaaaaca$				
25.5	E E G G D Y Y D D E L F S D V Q D I K T	MVP gene			
2767	gccttggccaaaatacacgaggataatcagaagataatctctaaactagaatcactgctg A L A K I H E D N Q K I I S K L E S L L	MVP gono			
2827	ttattgaagggagaagttgagtcaattaagaagcagattaacaggcaaaatatcagcata	MVP gene			
2027	L L K G E V E S I K K O I N R O N I S I	MVP gene			
2887	tccaccttggaaggacacctctcaagcatcatgatcgccattcctggacttgggaaggat	,			
	STLEGHLSSIMIAIPGLGKD	MVP gene			
2947	cccaacgaccccactgcagatgtcgaactcaatcccgacttgaaacccatcataggcaga				
	P N D P T A D V E L N P D L K P I I G R	MVP gene			
3007	gattcaggccgagcactggccgaagttctcaagaaacccgctgccagccgacaactccaa D S G R A L A E V L K K P A A S R O L O	MVP gene			
3067	ggaatgacaaatggacggaccagttccagaggacagctgctgaaggaattccaactaaag	nvr gene			
3007	G M T N G R T S S R G Q L L K E F Q L K	MVP gene			
3127	ccgatcgggaaaaaggtgagctcagccgtcgggtttgtccctgacaccgggcctgtatca	,			
	PIGKKVSSAVGFVPDTGPVS	MVP gene			
3187	cgcagtgtaatccgctccattataaaatccagtcggctagaagaggatcggaagcgttac				
20:-	R S V I R S I I K S S R L E E D R K R Y	MVP gene			
3247	ctgatgactctccttgatgatatcaaaggagccaacgatcttgccaagttccaccagatg	MVP gono			
3307	L M T L L D D I K G A N D L A K F H Q M ctgatgaaqataataatgaagtag	MVP gene			
3307	L M K I I M K -	MVP gene			
Figure 1. Sequence comparison of the open reading frames of Measles virus P/V/C genes. The shared ATG start of the MVP/MVV genes is					

Figure 1. Sequence comparison of the open reading frames of Measles virus P/V/C genes. The shared ATG start of the MVP/MVV genes is shown in red. The shared amino acid sequence of MVP/MVV is shown in black. The unique amino acid sequence of MVV, caused by a non-templated G insertion into the sequence at nucleotide 2499 (site of insertion is boxed in red, inserted base not shown), is shown in red. The start ATG for MVC and the unique amino acid sequence are shown in blue. Sequence from GenBank NC_001498.1.

Key Research Accomplishments

Nothing to report

Conclusion

This work has the potential to significantly alter our understanding of Paget's disease. For the past 37 years, the role of Measles virus in Paget's disease has been a highly controversial question. Our model has the potential to address this question directly and to provide a model for the role of Measles virus and how Paget's disease initiates.

As noted, we have requested a no-cost extension of the grant to complete the work. We have identified several alternative strategies, which are outlined in the Overall Project Summary, to overcome the problems that we have encountered in the experimental design of both aims. With the additional time requested in the no-cost extension, we will be able to complete the objectives of the grant.

Publications, Abstracts and Presentations

Nothing to report

Inventions, Patents and Licenses

Nothing to report

Reportable Outcomes

Nothing to report

Other Achievements

Nothing to report

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Appendices

Nothing to report